

Amendments to the Specification:

Please replace the paragraph on page 26, lines 1-29, with following rewritten paragraph:

A number of fucosyltransferases are known to those of skill in the art. Briefly, fucosyltransferases include any of those enzymes, which transfer L-fucose from GDP-fucose to a hydroxy position of an acceptor sugar. In some embodiments, for example, the acceptor sugar is a GlcNAc in a Gal β (1 \rightarrow 3,4)GlcNAc group in an oligosaccharide glycoside. Suitable fucosyltransferases for this reaction include the known Gal β (1 \rightarrow 3,4)GlcNAc α (1 \rightarrow 3,4)fucosyltransferase (FucT-III E.C. No. 2.4.1.65) which is obtained from human milk (see, e.g., Palcic et al., *Carbohydrate Res.* 190:1-11 (1989); Prieels, et al., *J. Biol. Chem.* 256:10456-10463 (1981); and Nunez, et al., *Can. J. Chem.* 59:2086-2095 (1981)) and the β Gal(1 \rightarrow 4) β GlcNAc α 1 \rightarrow 3)fucosyltransferases (FucT-IV, FucT-V, FucT-VI (MDPLGPAPKQWSWRCCLTTLFQLLMVCFFSYLRVSQDDPTVYPNGSRFPDSTGTAPAHSIPLILLWTWPFNKPIALPRCSEMVPGTADCNITADRKVYPQADAVIVHHREVMYNPSAQLPRSPRRQGGQRWIWFSMESPSHCWQLKAMDGYFNLTMSYRSDSIDFTPYGWLEPWSGQPAHPPLNLSAKTELVAWAVSNWGPNSARVRYQSLQAHLKVDVYGRSHKPLPQGTMMETLSRYKFYLAFENSLHPDYITEKLWRNALEAWVPVVLGSPRSNYERFLPPDAFIHVDDFQSPKDLARYLQELDKDHARYLSYFRWRETLRPRSFSWALAFCKACWKLQEESRYQTRGIAAWFT (SEQ ID NO:1)), and FucT-VII (MNNAGHGPTRRRLRGLVLAGVALLAALWLLWLLGSAPRGTPAPQPTITILVWHWPFTDQPELPSDTCTRYGIARCHLSANRSLASADA VVFHHRELQTRRSHLPLAQRPQGQPVWVWASMESPSHTHGLSHLRGIFNWVLSYRRSDIFVPYGRLEPHWGPSPPLPAKSRVAAWVVSNFQERQLRARLYRQLAPHLRVDVFGGRANRPLCASC LVPTVAQYRFYLSFENSQHRDYITEKFWRNALVAGTVPPVVLGPPPRATYEAFVPADAFVHVDDFGSARELA AFLTGMNESRYQRFFAWDRDLRVRLFTDWRERFCAICDRYPHPLRSQVYEDLEGWFQA (SEQ ID NO:2)), E.C. No. 2.4.1.65) which are found in human serum. A recombinant form of β Gal(1 \rightarrow 3,4) β GlcNAc α (1 \rightarrow 3,4)fucosyltransferase is also available (see, Dumas, et al., *Bioorg. Med. Letters* 1: 425-428 (1991) and Kukowska-Latallo, et al., *Genes and Development* 4: 1288-1303 (1990)). Other exemplary fucosyltransferases include α 1,2 fucosyltransferase (E.C. No. 2.4.1.69). Enzymatic fucosylation may be carried out by the methods described in Mollicone et al., *Eur. J. Biochem.* 191:169-176 (1990) or U.S. Pat. No. 5,374,655; an α 1,3-fucosyltransferase from *Schistosoma mansoni* (Trottein et al. (2000) *Mol. Biochem. Parasitol.* 107: 279-287); and an α 1,3 fucosyltransferase IX (nucleotide sequences of human and mouse FucT-IX are described in Kaneko et al. (1999) *FEBS Lett.* 452: 237-242, and the chromosomal location of the human gene is described in Kaneko et al. (1999) *Cytogenet. Cell Genet.* 86: 329-330. Recently reported α 1,3-fucosyltransferases that use an N-linked GlcNAc as an acceptor from the snail *Lymanaea stagnalis* and from mung bean are described in van Tetering et al. (1999) *FEBS Lett.* 461: 311-314 and Leiter et al. (1999) *J. Biol. Chem.* 274: 21830-21839, respectively. In addition, bacterial fucosyltransferases such as the α (1,3/4) fucosyltransferase of *Helicobacter pylori* as described in Rasko et al. (2000) *J. Biol. Chem.* 275:4988-94, as well as the α 1,2-fucosyltransferase of *H. pylori* (Wang et al. (1999) *Microbiology.* 145: 3245-53. See, also Staudacher, E. (1996) *Trends*

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in Glycoscience and Glycotechnology, 8: 391-408, for lists and descriptions of fucosyltransferases useful in the invention.